

Ecomicrobiological Study on River Nile at Kafr El-Zayat Industrial Area

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Abstract

The total microbial count including fungi, yeasts and total bacterial counts were recorded as indicators for faecal pollution in the River Nile (Rosetta branch) at Kafr El-Zayat Industrial Area. The highest counts for yeasts were recorded during winter in front of and at about 1 km far from El-Mallh and Soda Company (113×10^2 and 148×10^2 CFU ml⁻¹). The most common identified yeast organisms were *Saccharomyces cerevisiae* and *Kloeckera* spp. Fungi highest count was recorded during winter at El-Mallh and Soda Company at 37°C and represented by *Trichoderma* spp. *Candida albicans* and *Candida magnoliae* were the most abundant species during spring and summer at 27°C. The highest counts of total and faecal coliform bacteria were 11×10^3 CFU ml⁻¹ during summer and winter. Gram-negative bacteria (i.e. pathogenic bacteria) had the highest count during winter, where *Klebsiella pneumoniae*, *Serratia liquefaciens*, *Salmonella choleraesuis* and *Escherichia coli* were the most common isolated organisms. All microbiological parameters indicated that Rosetta Branch was subjected to sewage pollution. In addition, some environmental parameters such as temperature, conductivity and pH were monitored. The role of yeasts and bacteria in nitrogen cycle in the water revealed that a proposed waste water treatment unit could be beneficial before the discharge of the industrial wastes that will help in solving ammonia problem.

Keywords: River Nile, Rosetta Branch, Faecal pollution, Bacterial indicators, Pathogenic bacteria Yeasts, Fungi, Ammonification, Nitrification and Denitrification.

1. Introduction

The problem of the limited amount of water resources in Egypt acquires more attention to the problem of over population and the inability to implement the projects of increasing water resources of the River Nile (Ahmed *et al.*, 1995). The Nile is the main source of water for drinking, agriculture, industry, navigation, recreation and fish production. Thus, it is of dominating influence on the economic, cultural, public health, social and political aspects of the country (Mahrous, 1997 and Rabeh, 2007). Concerning the pollution of Rosetta Branch, there are three main sources which potentially affect and deteriorate its water quality. El-Rahawy drain pours more than 400,000 m³ daily including 398,000 m³ liquid sewage and 2,000 m³ of sludge from Giza Governorate (Ghallab, 2000). At the same time, Soble drain at Menofya Governorate pours its agricultural wastes directly into this branch. The second source is Kafr El-Zayat Industrial Area, which includes the industrial effluents from the factories of super-phosphate and sulfur-compounds (e.g. El-Malia Company), oil and soap industries (e.g. El-Mallh and Soda Company) and

pesticides factory, which pour their effluents directly into the branch without any treatment. The third source is many agricultural drains that discharge their waters into this branch, in addition to the sewage discharged from many cities and villages that are distributed along its two banks (Abdo, 2002).

The use of normal intestinal organisms as indicators of faecal pollution rather than the pathogens themselves is a universally accepted principle for monitoring and assessing microbial safety of water supplies. Organisms commonly used as bacterial indicators for faecal pollution are the coliform group including faecal coliforms and faecal streptococci (Anne-Marie *et al.* 1991; Hecter *et al.* 1998 and Rabeh, 2000). Total yeasts, especially *Candida albicans*, were identified and counted to determine the microbiological profile of River Nile at Cairo (EL-Taweel, 1998). Therefore, the present study aims at evaluating the ecomicrobiological characteristics of River Nile water at Kafr EL-Zayat Industrial Area (Rosetta Branch) through counting some yeasts (e.g. *Candida* spp and *Trichosporon* spp), fungi and total bacteria as microbial indicators in relation to some physico-chemical parameters.

2. Materials and Methods

2.1. Study Area

River Nile is considered as the longest river in the world. Its length is approximately 6740 km. Twenty five km North of Cairo at EL-Qanater Barrage, the River Nile bifurcates into two branches (Damietta and Rosetta). Rosetta Branch width varies from 150 to 200 m, with an average depth of 2 to 3.5 m (Abdo, 2002). The water level fluctuates between 10.6 and 13.8 meter above sea level (Sayed, 2002). Rosetta Branch begins from El-Qanater El-Khyria Barrage at the south and ends at the Rosetta Estuary in the Mediterranean Sea at the north. It is of great vital importance as a source for municipal, industrial and agricultural water, and for navigation and fish farms.

2.2. Sampling

Sampling process was carried out seasonally (November 2005 - August 2006). Five stations were selected for the collection of water samples from Rosetta Branch at Kafr El-Zayat (Figure 2): station 1 in front of El-Mallh and Soda Company, station 2 in front of El-Malia Company, station 3 in front of Benover Village, station 4 at about 1 km downstream El-Malia Company and station 5 at about 1 km downstream El-Mallh and Soda Company. For pesticides determination, water samples were collected in front of El-Malia Company during winter 2006 as it produces super-phosphate and sulfur-compounds and its effluents might include chemicals used in the production of pesticides.

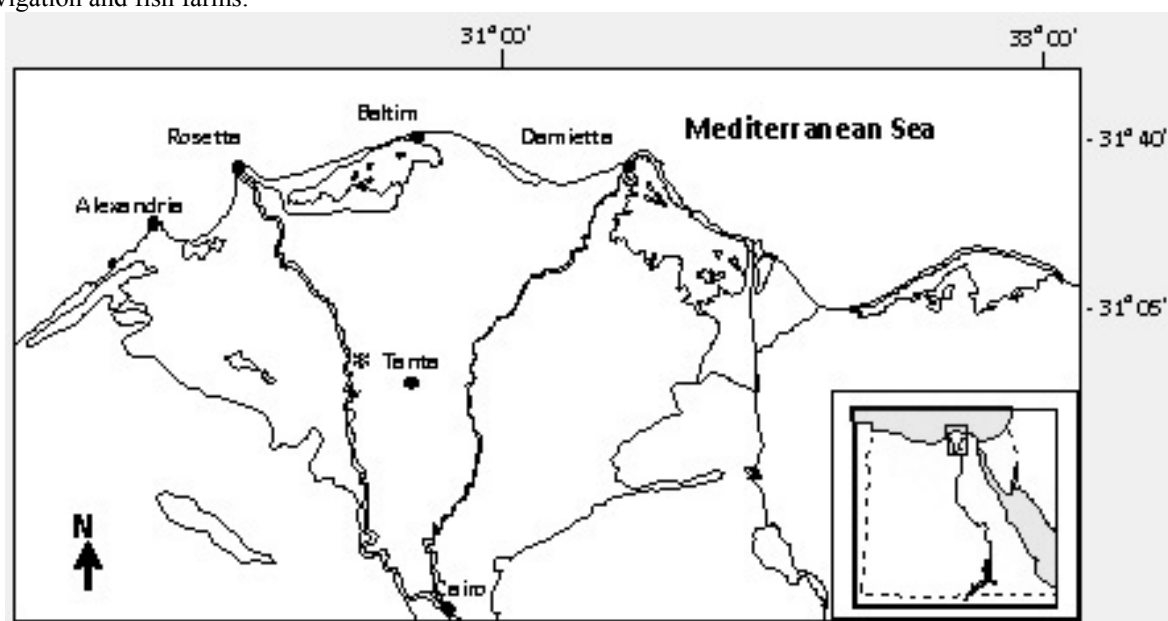


Figure 1. Map of Egypt indicates Kafr El-Zayat Industrial Area close to Rosetta Branch (*).



Figure 2: Sampling sites in the front of El-Malia (left side) and Mallh & Soda Companies (right side) at Kafr EL-Zayat Industrial Area.

2.3. Physical and chemical water analysis

Water sampling for chemical analysis were collected using plastic bottles. The samples were stored, refrigerated and analyzed immediately after arrival to the laboratory. For analysis of heavy metals, samples were preserved immediately after collection by acidifying with concentrated HNO₃ to pH < 2 using 5 ml nitric acid for 1 liter sample, then the samples were stored in a refrigerator at 4°C or in a deep freezer to prevent change in volume due to evaporation. Samples might be filtered if necessary, and a portion of the filtered sample was taken for the required determination. For pesticides analysis, samples were collected using plastic bottles (about 2 liters) and stored in a refrigerator at 4 °C. Air and water temperatures (°C), electrical conductivity (μS cm⁻¹), hydrogen ion concentrations (pH values) and salinity (EC) of water were measured directly at the sampling stations using water quality checker U-10 HORIBA. Dissolved oxygen was carried out using the modified Winkler method, Ammonia using Nesslerization method, Nitrate-Nitrogen using ultraviolet spectrophotometric screening method, and some heavy metals (Lead, Zinc, Copper, and Cadmium) using nitric acid digestion method (APHA, 1998). Some pesticides were measured using Gas chromatography (GC/ECD) according to UNEP/IOC/IAEA (1991) and IOC (1993).

2.4. Microbiological analysis

Sterile 100 ml plastic bottles were used in this analysis (Rabeh, 1993). Surface plate method (Rabeh, 1996) was used for determination of the total bacterial count at 22°C for 72 h and at 37°C for 48 h using Plate Count Agar medium. Most probable number method (MPN) was used for determination of total coliform (TC) and faecal coliform (FC) on MacConkey Broth (at 37°C for 48 h and 44.5°C for 24 h, respectively). Azide dextrose broth was used for determining MPN of faecal streptococci (FS) at 37°C for 48 h (APHA, 1998). Gram negative bacteria were enumerated in the examined water on MacConky agar medium using pour plate method (see Rabeh and Azab, 2006). All isolated Gram-negative bacteria were identified using API 20E (bioMerieux). From each plate containing specific medium, one colony was added to 5ml sterile water. The solution was inoculated into the strip, containing mini-test tubes, and incubated for 18-24 h at 37°C. The coloration reactions were read and the reaction converted to a digital code which compared to the Reading table provided by bioMerieux.

The plate count technique was used for the isolation and enumeration of fungi (Dutka, 1989). Using Aureomycin Rose Bengal Glucose Peptone Agar (ARGPA), 0.1 to 1 ml of the sample was plated out, and left to be dried. After drying, the Petri dishes were inverted and incubated aerobically at 27°C for 3 days. Incubation was also carried out at 37°C for 1 or 2 days, and then the number of colonies were counted on each

plate and recorded as CFU ml⁻¹. After enumeration, fungal colonies were transferred to Peptone Dextrose Agar or Sabouraud's Dextrose Agar plates for identification purposes. The identification process was according to Barnett (1972) for the genera of fungi imperfecti, Barron (1968) for the genera of *Hyphomycetes*, Domsch *et al.* (1980) for compenedium of soil fungi, Ramirez (1982) for *Penicillia* and Raper and Fennell (1965) for *Aspergillus* species. The plate count technique was used for the isolation and enumeration of yeast (Dutka 1989). Suspected colonies of yeast were streaked onto sabouraud dextrose agar plates, and incubated for 48 h at 28°C. These cultures were used as the source of inoculum for the yeast confirmatory tests. The isolated yeasts were identified according to Lârone (1995) using API 20 CAUX from Temco Technology Company (Cairo).

Determination of the MPN of ammonifying and nitrifying microorganisms was carried out and calculated using tables of Cochran (1950). The culture nitrite and nitrate were determined after 21 days incubation by diphenylamine indicator. Also, MPN method was used for counting nitrate-reducing bacteria. The culture nitrate was tested on nitrate-reducing medium after 14 days incubation using sulfanilic acid and Naphthylamine solutions as indicators for the presence of nitrite (Naguib, 1961). Ammonification, nitrification and nitrate-reduction processes were carried out according to Rheinheimer (1959). Rate of ammonia production was determined using 5 ml of media for yeast (consisting of 10 g asparagine and 40 g glucose dissolved in 1 liter distilled water) with 50 ml of water sample. After 4, 8, 12 and 15 days of incubation at 25°C, the amount of the released NH₄-N was determined calorimetrically using Nessler's reagent (APHA, 1998). These processes were controlled by the same amount of the medium with the addition of 50 ml of the examined water, previously sterilized in autoclave and distilled water, in order to make sure that the NH₄-N at the end of the experiment is not produced as a result of atmospheric diffusion.

3. Results

The highest water temperature was recorded during summer (30.3°C), while the lowest was during autumn (19.4°C). The maximum electrical conductivity (EC) was recorded in front of El-Malia Company (723 μS cm⁻¹), while the minimum was at about 1 km downstream of El-Malh and Soda Company (559 μS cm⁻¹). The maximum value of dissolved oxygen was recorded in front of El-Malia Company (4.8 mg l⁻¹), while the minimum was in front of Benover Village (3.8 mg l⁻¹). The results showed that COD fluctuated between 11.2 in winter and 20.8 mg l⁻¹ in summer (Tables 1 and 2). Generally, estimation of NH₃-N values indicated high values during winter. On the other hand the maximum value of nitrite was recorded during spring (0.35 μg l⁻¹). The average values of

nitrate varied from 4.65 to 6.07 $\mu\text{g l}^{-1}$ between seasons, and from 5.4 to 4.9 $\mu\text{g l}^{-1}$ between stations. The maximum value of copper was recorded in front of Benover Village during spring (0.04 ppm), while the maximum of lead (0.33 ppm) was recorded at about 1 km downstream El-Malia Company. The values recorded for zinc were 0.13, 0.17, 0.02 and 0.06 ppm, while those of cadmium were 0.10, 0.19, 0.01 and 0.06 ppm in autumn, winter, spring and summer, respectively. The total concentration of all measured pesticides was 316.4 ng l^{-1} .

3.1. Counts of total (TBC) and Gram-negative bacteria (G-N)

The highest values of TBC at 22°C (600 $\times 10^3$ CFU ml^{-1}) and at 37°C (1000 $\times 10^3$ CFU ml^{-1}) were recorded during summer (Table 3). On the other hand, the lowest TBC at 22°C was recorded in front of Benover Village and at about 1 km downstream El-Malia Company during summer and autumn (14 $\times 10^3$ CFU ml^{-1}), while the lowest at 37°C was recorded in front of El-Malia Company during spring (2 $\times 10^3$ CFU ml^{-1}). The highest value of G-N was recorded in front of El-Malah and Soda Company during winter (31000 $\times 10^2$ ml^{-1}). Four isolates, namely *Escherichia coli*, *Salmonella choleraesuis*, *Serratia liquefaciens* and *Klebsiella pneumoniae*, were commonly found in all stations during the four seasons.

Table (1): Seasonal variation of the physico-chemical characters (mean \pm SD) of the River Nile water at Kafr El-Zayat Industrial Area during 2005-2006. Seasonal maximum values are underlined.

Character	2005 Autumn	2006 Winter	2006 Spring	2006 Summer	
Temperature ($^{\circ}\text{C}$)	19.40 \pm 0.40	19.60 \pm 1.00	24.90 \pm 0.40	<u>30.30 \pm 1.30</u>	
EC ($\mu\text{S cm}^{-1}$)	<u>752.00 \pm 94.00</u>	681.00 \pm 45.00	487.00 \pm 33.00	598.00 \pm 138.00	
pH	7.80 \pm 0.25	7.60 \pm 0.10	7.40 \pm 0.08	7.80 \pm 0.13	
Salinity (%)	<u>0.03 \pm 0.01</u>	0.02 \pm 0.01	0.01 \pm 0.01	0.02 \pm 0.01	
D Oxygen (mg l^{-1})	3.20 \pm 0.60	2.10 \pm 0.50	4.00 \pm 0.60	<u>7.00 \pm 0.70</u>	
COD (mg l^{-1})	19.20 \pm 11.50	11.20 \pm 5.20	14.40 \pm 7.30	<u>20.80 \pm 6.60</u>	
Ammonia	2.80 \pm 0.05	<u>3.60 \pm 0.28</u>	0.80 \pm 0.10	0.30 \pm 0.18	
Nitrite	$\mu\text{g l}^{-1}$	0.12 \pm 0.02	0.13 \pm 0.01	0.21 \pm 0.01	
Nitrate		4.65 \pm 0.93	4.92 \pm 0.44	<u>6.07 \pm 0.23</u>	4.87 \pm 0.87
Cu ⁺³		0.01 \pm 0.01	0.03 \pm 0.02	<u>0.07 \pm 0.05</u>	0.01 \pm 0.01
Pb ⁺²	ppm	0.06 \pm 0.06	0.19 \pm 0.27	<u>0.41 \pm 0.39</u>	0.13 \pm 0.09
Zn ⁺		0.13 \pm 0.09	<u>0.17 \pm 0.34</u>	0.02 \pm 0.05	0.06 \pm 0.06
Cd ⁺³		0.10 \pm 0.09	<u>0.19 \pm 0.39</u>	0.01 \pm 0.01	0.06 \pm 0.05

Table (2): Spatial variation of the physico-chemical characters (mean \pm SD) of the River Nile water at Kafr El-Zayat Industrial Area during 2005-2006. Station 1: in front of El-Mallh & Soda Company, Station 2: in front of El-Malia Company, Station 3: in front of Benover Village, Station 4: at about 1 km downstream of El-Malia Company, Station 5: at about 1 km downstream of El-Mallh & Soda Company. Seasonal maximum values are underlined.

Character	Station 1	Station 2	Station 3	Station 4	Station 5	
Temperature ($^{\circ}\text{C}$)	24.50 \pm 6.00	<u>23.80 \pm 5.00</u>	23.10 \pm 5.00	23.10 \pm 5.00	23.20 \pm 5.00	
EC ($\mu\text{S cm}^{-1}$)	686.00 \pm 118.0	<u>723.00 \pm 144.00</u>	614.00 \pm 152.00	566.00 \pm 104.0	559.00 \pm 95.0	
pH	7.70 \pm 0.28	7.70 \pm 0.18	7.60 \pm 0.28	7.70 \pm 0.24	7.60 \pm 0.19	
Salinity (%)	0.02 \pm 0.01	0.02 \pm 0.01	0.02 \pm 0.01	0.01 \pm 0.01	0.01 \pm 0.01	
D Oxygen (mg l^{-1})	4.40 \pm 2.00	<u>4.80 \pm 2.30</u>	3.80 \pm 2.40	3.90 \pm 1.90	4.40 \pm 2.10	
COD (mg l^{-1})	17.00 \pm 7.00	13.00 \pm 5.00	19.00 \pm 12.00	11.00 \pm 9.00	<u>22.00 \pm 5.00</u>	
Ammonia	1.70 \pm 1.70	1.80 \pm 1.40	1.90 \pm 1.60	1.90 \pm 1.50	1.80 \pm 1.40	
Nitrite	$\mu\text{g l}^{-1}$	0.20 \pm 0.10	0.20 \pm 0.10	0.20 \pm 0.10	0.20 \pm 0.10	
Nitrate		5.30 \pm 0.90	5.20 \pm 0.60	<u>5.40 \pm 1.00</u>	4.90 \pm 1.00	4.60 \pm 0.70
Cu ⁺³		0.03 \pm 0.04	0.02 \pm 0.03	<u>0.04 \pm 0.04</u>	0.03 \pm 0.03	0.02 \pm 0.02
Pb ⁺²	ppm	0.25 \pm 0.20	0.05 \pm 0.10	0.24 \pm 0.30	<u>0.33 \pm 0.40</u>	0.15 \pm 0.10
Zn ⁺		0.07 \pm 0.06	<u>0.26 \pm 0.35</u>	0.07 \pm 0.11	0.02 \pm 0.03	0.06 \pm 0.09
Cd ⁺³		0.03 \pm 0.05	0.06 \pm 0.07	0.06 \pm 0.09	<u>0.24 \pm 0.43</u>	0.04 \pm 0.08

Table 3: Total bacterial counts (CFU × 10³ ml⁻¹), total count of Gram-negative bacteria (G-N × 10².100 ml⁻¹) of the River Nile water at Kafr EL-Zayat Industrial Area during 2005-2006. Station 1: in front of El-Mallh & Soda Company, Station 2: in front of El-Malia Company, Station 3: in front of Benover Village, Station 4: at about 1 km downstream of El-Malia Company, Station 5: at about 1 km downstream of El-Mallh & Soda Company. G-N: Gram-negative bacteria.

S	Autumn 2005			Winter 2006			Spring 2006			Summer 2006		
	22°C	37°C	G-N	22°C	37°C	G-N	22°C	37°C	G-N	22°C	37°C	G-N
1	400	276	1000	164	172	31000	128	232	3000	600	1000	4700
2	80	31	1000	56	180	13000	28	2	5000	84	4	4200
3	172	141	0.0	36	56	100	56	72	3000	14	60	1600
4	14	44	1000	164	400	17000	96	32	1000	23	36	3000
5	17	12	0.0	28	108	0.0	22	4	21000	22	17	1900

3.2. Total coliforms (TC), faecal coliforms (FC) and Faecal streptococci (FS)

The highest counts of TC were recorded in most stations during summer and winter (11000 CFU.100 ml⁻¹), while the lowest was recorded in spring (90 CFU 100 ml⁻¹). FC counts fluctuated between nil in station 3 during autumn and 11000 CFU.100 ml⁻¹ in front of El-Malia Company during summer. Compared with the other stations, the highest counts of FC were recorded in front of El-Malia Company during summer and winter (11000 and 4600 CFU 100 ml⁻¹). FS varied from 150 during winter to 110000 CFU 100.ml⁻¹ during autumn and spring (Table 4).

3.3. Yeast and fungal species as microbial indicator of water pollution

Highest count of yeasts (148x10² CFU ml⁻¹) was recorded during winter at 37°C at about 1 km downstream of El-Malah & Soda Company (Tables 5). Most of the isolated yeasts were *Kloeckera* spp and *Saccharomyces cerevisiae*. On the other hand, the highest count of fungi (15x10² CFU ml⁻¹) during winter at 37°C in front of El-Mallh & Soda Company (*Trichoderma* spp was the most abundant). The isolated yeasts were *Rhodotorula glutinis*, *Cryptococcus laurentii*, *Kloeckera* spp, *Saccharomyces cerevisiae*, *Candida albicans*, *Candida magnoliae*, *Trichosporon*

mucooides and *Candida glabrata*. The most common identified fungi were *Aureobasidium pullulans*, *Cladosporium carinii*, *Asperigillus tamari*, *Xylohypha bantiana*, *Trichoderma* spp, *Blastomyces* spp and *Penicillium mareiffei*.

3.4. Bacteria involved in nitrogen cycle

Ammonifying, nitrifying and denitrifying bacteria in River Nile water were counted during winter in front of Benover Village (it had the highest ammonia concentration during 2006). The most probable number of ammonifying bacteria was 240 colony ml⁻¹, and that of nitrifying bacteria was 39 colony ml⁻¹ during winter. On the other hand, denitrifying bacteria were completely absent from the studied station. High value of 148 µg l⁻¹ of water ammonification process was measured (Table 6).

3.5. Role of Yeasts in Nitrogen Cycle:

The result of the laboratory experiment on ammonification process by yeasts, which inhabiting River Nile water after different incubation periods, indicated that the value of ammonification process was higher, after 4 days of incubation, in non-autoclaved water sample (28.2 µg l⁻¹) than in autoclaved water sample, 2.9 µg l⁻¹ (Table 7).

Table 4: Most probable number (MPN) of bacterial indicators of sewage pollution (CFU.100 ml⁻¹) of the River Nile water at Kafr EL-Zayat Industrial Area during 2005-2006. Station 1: in front of El-Mallh & Soda Company, Station 2: in front of El-Malia Company, Station 3: in front of Benover Village, Station 4: at about 1 km downstream of El-Malia Company and Station 5: at about 1 km downstream of El-Mallh & Soda Company.

S	Autumn 2005			Winter 2006			Spring 2006			Summer 2006		
	TC	FC	FS	TC	FC	FS	TC	FC	FS	TC	FC	FS
1	3900	3900	110000	4600	4600	2300	4600	230	46000	2300	2300	93000
2	2100	2100	1500	11000	4600	2300	230	230	110000	11000	11000	4300
3	400	0.00	7500	11000	930	7500	90	90	46000	11000	200	46000
4	11000	90	4300	430	230	3900	430	90	15000	2400	70	21000
5	1200	230	3900	430	230	150	150	90	9300	2100	210	15000

Table 5: Total yeast and fungi counts at 27 °C and 37 °C ($\times 10^2 \text{ ml}^{-1}$) of the River Nile water at Kafr EL-Zayat industrial area during 2005- 2006. Station 1: in front of El-Mallh & Soda Company, Station 2: in front of El-Malia Company, Station 3: in front of Benover Village, Station 4: at about 1 km downstream of El-Malia Company, Station 5: at about 1 km downstream of El-Mallh & Soda Company.

Season	Autumn 2005				Winter 2006				Spring 2006				Summer 2006			
	Yeast count		Fungal count		Yeast count		Fungal count		Yeast count		Fungal count		Yeast count		Fungal count	
	27	37	27	37	27	37	27	37	27	37	27	37	27	37	27	37
°C																
1	2	1	2	0	113	128	3	15	7	30	1	0	3	2	0	0
2	1	1	0	1	0	24	1	0	2	9	0	0	0	1	0	0
3	0	0	2	2	97	29	0	0	5	10	0	1	2	0	1	1
4	0	0	0	0	2	143	1	0	4	1	1	0	0	1	0	5
5	0	1	0	0	0	148	0	1	9	2	0	0	1	1	0	0

Table 6: Ammonification, nitrification and denitrification processes ($\mu\text{g l}^{-1}$) in the River Nile water at Kafr El-Zayat Industrial Area during winter 2006. C: autoclaved water sample, S: non-autoclaved water sample, ND: not detected.

Process	Ammonification		1 st step of Nitrification		2 nd step of Nitrification		Denitrification	
	C	S	C	S	C	S	C	S
NH ₃	9.30	148	ND	ND	ND	ND	2.15	0.90
NO ₂	ND	ND	0.16	0.93	ND	ND	0.18	0.15
NO ₃	ND	ND	ND	ND	24.98	25.07	ND	ND

Table 7: Ammonification processes ($\mu\text{g l}^{-1}$) by yeasts inhabiting the River Nile water at Kafr El-Zayat Industrial Area during winter 2006.

Time (day)	Autoclaved water sample	Non-autoclaved water sample
4	2.9	28.2
8	3.9	24.6
12	1.7	12.5
15	0.9	13.5

4. Discussion

Variety of wastes effluent discharge, either without or with partial treatment into the Nile, resulting in changes in its physical and chemical conditions and microbial quality. Water temperature is an important factor which affects the biological activities of aquatic organisms (Bojanic *et al.* 2001), and the amount of oxygen that can be dissolved in water. In the present study, elevation of temperature in front of EL-Mallh and Soda Company all over the year could be attributed to the discharge of hot waste water into the Nile, which consequently induces thermal pollution leading to the discharge of hot waste water into the Nile, which consequently induces thermal pollution leading to elevation of temperature. This finding agrees with Bedair (2006). In addition, the high value of EC in the Nile water may be attributed to the discharge of El-Rahawy drain which pours more than 400,000 m³ daily including 398,000 m³ liquid sewage and 2,000 m³ of sludge from Giza Governorate. Soble drain pours a great amount of agricultural effluents containing large amounts of dissolved ions and high amount of organic

and inorganic constituents. This was in accordance with the study of Elewa and Mahdi (1988). The present results indicated also an obvious increase in the water salinity during autumn and summer. The increase in water salinity in summer may be due to the increase in temperature and consequently increasing evaporation (Rabeh 1993). pH of the River Nile water is always in the alkaline side with slight variation between different stations (El-Bassat 1995 and 2002, Heikal 2000, Abdo 2002 and Bedair 2006).

Dissolved oxygen (DO) is the best parameter to show the effect of pollution on a water system unless it contains toxic compounds. Oxygen is also needed for all oxidation, nitrification and decomposition processes and is controlled by three factors: photosynthesis, respiration and exchange at the air-water interface (Krom *et al.* 1989, Erez *et al.* 1990). In the present investigation, values of dissolved oxygen ranged between 1.5 mg l⁻¹ during winter in the front of Benover village and 8.1 mg l⁻¹ during summer in the front of El-Malia Company. Generally, DO concentrations in the Nile are always higher than 7.0 mg l⁻¹, indicating high assimilation capacity (Ezzat *et al.* 2002). According to APHA (1998), the chemical

oxygen demand (COD) is used as a measure of the oxygen equivalent to organic matter content of a sample that is susceptible to oxidation by a strong chemical oxidant. The highest COD value ($36 \text{ mg O}_2 \text{ l}^{-1}$) was recorded in the front of Benover village during autumn. This might be due to low water flow which reflects the high load of organic matter at this station. Generally, during winter the COD value was higher at some stations, this may be due to the winter closure which started by the end of December causing decrease in the level of Nile water (this leads to elevate the organic load of the Nile during this period). Also the relative increase of COD values during summer season may be attributed to the increase in water temperature which facilitates the decomposition of organic matter in water that leads to increase COD values (Ali, 1998).

Reid (1961) reported that ammonia-nitrogen in excess of 1 mg l^{-1} has been given as an indicator of organic pollution and can be toxic to aquatic species in concentration over 2.5 mg l^{-1} . Higher concentration of ammonia in the area of the present study could be attributed to organic pollution resulted from domestic sewage, industrial waste and fertilizer runoff. The high values of NH_3 recorded during hot seasons especially in spring (ranged between 0.63 and 0.91 mg l^{-1}) may be due to high evaporation rates depending on the elevated temperature, in addition to the denitrification process by the reduction of NO_2 and NO_3 to NH_3 under lower values of pH and DO (Elewa *et al.* 2001). The pronounced increase in NH_3 content during winter, especially in the front of Benover village, may be attributed to the decomposed organic matter and dying off of algal blooms resulted in increasing of ammonia concentration (Krom *et al.* 1989).

During winter and autumn, NH_3 had positive correlations with NO_3 , total count of yeasts and total coliform.. This indicates that high concentration of ammonia was correlated with high level of faecal pollution. Generally, surface water not influenced by human activities contains nitrate concentration less than $5 \mu\text{g l}^{-1}$. Excess of $5 \mu\text{g l}^{-1}$ usually indicates pollution by human or animal waste or/and fertilizers run off (Chapman 1992). Nitrate is generally considered the only thermodynamically stable form of nitrogenous compounds in absence of oxygen (Horna 1972). In case of nitrate, the highest value was recorded during summer ($6.41 \mu\text{g l}^{-1}$). This agrees with Heikal (2000) who found that nitrate value ranged between 3.5 - $6.8 \mu\text{g l}^{-1}$ in River Nile water. Most of the maximum values of NO_3 were recorded in the front of El Mallh & Soda and El-Malia Companies which mainly attributed to the effects of agricultural and industrial effluents poured close to these stations. Comparable results were reported by Siliem (1984). On the contrary, the relative decrease in NO_3 - contents recorded during autumn may be attributed to the drought period in addition to the denitrification of NO_3 into NO_2 and NH_3 by denitrifying bacteria.

Nitrite is an intermediate oxidation state of nitrogen, both in the oxidation of ammonia to nitrate

and in the reduction of nitrate. The relative increase of NO_2 concentrations during summer and spring may be due to the increase in temperature and the oxidation of ammonia, while the sharp decrease during autumn may be due to the increase of the oxidation rate of nitrite to nitrate. Generally, nitrite is present in water as an ion or free acid and both are toxic (Russo *et al.*, 1981). NO_2 concentrations during summer and spring were high at the front of El-Mallh & Soda Company and this may be attributed to the industrial effluent from this factory, while the lowest value (in most seasons) was recorded at about 1 km downstream El-Malia and El-Mallh & Soda Companies.

Heavy metals have environmental persistence, almost of them have no toxicity at low concentration, but they have ability to incorporate into food chain and thus it is possible to be concentrated to toxic levels by the aquatic organisms. They are usually divided into two subclasses: Co, Cu, Fe, Mn and Zn which are essential for the correct functioning of biochemical processes; and Hg, Cd, Cr, and Pb that have no established biological function and represent the most important contaminants in the aquatic environment. In the study area, during autumn, the relative decrease of copper concentration may be attributed to its adsorption by humic materials that deposited to bottom, in addition to dilution effect of flood period (Elewa, 1991). On the other hand, the high values of copper contents were recorded during spring and summer, this may be due to the high evaporation rate under high temperature of water and air during hot seasons, and the release of Cu from sediment to the overlying water (Abdo, 1998). The high average values of Cd content in the study area were observed during winter, while the low values were during spring and summer. Comparable results were reported by (Bedair, 2006).

DDT compounds are used in fighting insects, mosquitoes and flies in the environment; they are converted to DDE compound by the effect of metabolic processes. Concerning the present study, the concentration of DDT compound was higher than that of DDE. This indicated that the DDT was recently present compared with DDE, which may be due to the discharge of El-Malia Company (it produces superphosphate and sulfur-compounds that include chemicals used in the production of pesticides). However, the total concentration of all measured pesticides in the area of investigation was 316.4 ng.l^{-1} . This value was within the permissible limits ($2 \times 10^6 \text{ ng.l}^{-1}$) according to the Egyptian Environmental Law no. 4 in 1994.

Total bacteria developing at 20 - 22°C are saprophytic types and non-pathogenic to human beings, while those developing at 37°C are mainly or potentially parasitic types derived from the soil, sewage, or excretal materials (APHA, 1998). The total count of bacteria at 22 and 37°C in the present study were high in summer and began to decrease in autumn at 22°C . This pattern was coincided with water temperature during the same seasons. The same

observations were previously reported by Rabeh (2007) and Rabeh *et al.* (2007). However, Abo-Sedra (1990) reported that the effect of temperature seems to be of little influence on the question of bacterial changes in River Nile waters. In Rosetta Branch, Rabeh (2007) reported that the highest bacterial counts (66.8×10^7 and 65.6×10^7 CFU ml⁻¹) at both incubations temperatures (22 and 37°C) were recorded during summer. On the other hand, the highest counts were recorded at Sobol Drain in Menofya Governorate, while the lowest were at Rosetta Estuary (0.3×10^7 and 0.2×10^7 CFU ml⁻¹).

The ratio of colony counts at the two incubation temperatures (22 and 37°C) helps to explain any sudden fluctuation in the bacterial count. This ratio is usually more to one in non-polluted waters, while in polluted waters it is below one. It was noticed that the ratio of bacteria recovered at 22 and 37°C was almost constant in all the examined water samples. In other words, an increase or decrease in the counts of bacteria developing at 22°C was associated with a similar trend to those developing at 37°C. The ratio of the count at 22°C to that at 37°C was slightly higher than 1, except in winter (0.51). So, the investigated area of the River Nile would be considered heavily polluted with sewage (APHA, 1998).

Total and faecal coliforms and faecal streptococci are indicators that have been used for decades to infer the presence of other potentially harmful pathogens in water (Noble *et al.*, 2004, Rabeh, 2007 and Sabae and Rabeh, 2007). The highest value of most probable number of coliform bacteria was recorded during summer in the front of El-Malia Company, which discharges nitrogenous wastes that may encourage the growth of indicator bacteria, in addition to the sewage wastes in River Nile water which also cause the high count of these indicator bacteria in the front of Benover Village. This result agrees with that reported by Rabeh (2007). On the other hand, total coliform count increased also during winter at the same stations, which may be due to the low flow period that leads to high concentration of organic matter, sewage and industrial wastes. Also high positive correlation between NO₃ and most of faecal indicator parameters might be due to the use of NO₃ by these organisms as nitrogen sources.

Several investigators prefer faecal streptococci (FS) instead of the coliforms in determining the level of sewage pollution (Dutka, 1989, Dan and Koppel, 1992). During this investigation, FS counts were higher during autumn than spring in the front of El-Mallh & Soda and El-Malia Companies as they discharge industrial wastes contains, super-phosphate, nitrogenous and sulfur compounds. These discharges encourage the bacterial growth. FS counts not only have their own indication to pollution, but also their ratio to faecal coliforms (FC) points to the source of faeces whether it is human (> 4) or animal (< 0.7) (Geldreich 1974). Accordingly, the FC/FS ratio in the present study indicates human pollution (> 4) in winter

and summer and of mixed source during autumn (3.0), while it was from animal source during spring (0.05). There is a positive correlation in the present study during spring between the two types of enteric bacteria, faecal coliform and faecal streptococci which supports the view that coliform and faecal streptococci are useful indicators for the presence of entero pathogens.

Bacterial indicators are often associated with disease-causing microorganisms of importance to public health (Godfree *et al.*, 1997) as these pathogens can lead to water-borne or water-borne related outbreaks (Martinez-Manzanares *et al.*, 1991). The highest total Gram-negative bacterial counts were recorded during winter, while the lowest were in autumn. This might be due to the high load of industrial, agricultural, human activities, and recreational activities which were discharged directly into the Nile. Comparison of the results obtained for characterization of our four isolates with those described in Holt *et al.* (1994), the isolates of the present study could be identified as *Escherichia coli*, *Salmonella choleraesuis*, *Serratia liquefaciens* and *Klebsiella pneumoniae*. *Candida albicans* is a known component in the humans and animals bodies, commonly found in the feces of humans and animals, thus it is an indicator of faecal contamination (Dutka 1989). Buck (1977) noted that *Candida albicans* was not found in treated sewage effluent. Shaban and El-Taweel (2003) counted *Candida albicans* and total yeast as indicators of fecal pollution in River Nile. Concerning yeast species contamination to water, Wójcik *et al.* (2003) revealed 28 yeast species representing seven genera: *Candida*, *Cryptococcus*, *Geotrichum*, *Kloeckera*, *Rhodotorula*, *Saccharomyces* and *Trichosporon* in the littoral zone of the Sulejów Reservoir of Poland. Uden and Ahearn (1963) had recorded many species of yeasts such as *Cryptococcus laurentii*, *Rhodotorula glutinis*, *Trichosporon mucoides* and *Cryptococcus albidusi* which were related to their founding species. *Aspergillus tamarii* and *Penicillium mareiffei* were recorded at different seasons during the period of investigation; these species were also appeared in the River Nile water near Sohag (El-Sharouny, 1989). Kiziewicz (2004) in his study on *Candida albicans* and *Candida tropicalis* found that their occurrence was associated with high concentration of ammonium nitrogen and suspended substance.

High rate of ammonification during winter, as indicated in the present study, may be the reason for ammonia accumulation resulting in death of fish. High count of ammonifying bacteria was accompanied with low count of denitrifying bacteria (see Jørgensen, 1989). Nitrifying bacteria is another important group of bacteria engaged in nitrogen cycle, which oxidised reduced nitrogenous matter to nitrite and then to nitrate (Hesselsøe, 2001), and make link between reduced and oxidized nitrogen compounds (Prinčič *et al.*, 1998). Variation in the number of nitrite and nitrate bacteria usually correspond to variations in the rate of NH₄-N and NO₂-N oxidation (Niewolak *et al.*, 1979). The

present study indicated that River Nile at Kafr El-Zayat industrial area is considered a highly contaminated one due to the presence of high numbers of indicator bacteria and yeasts. Furthermore it demonstrated a difference in the distribution of bacteria involved in nitrogen cycle which played a significant role in nitrogen removal, promoted ammoniation, nitrification and denitrification in river. In conclusion, the present study suggests pretreatment of wastewater through microbiological unit before the discharge of industrial wastes of El-Malia and El-Mallh & Soda Companies which may solve the accumulation of NH_3 , that leads to fish death in Nile.

References

- Abdo, M.H.: 1998, Some environmental studies on the River Nile and Ismailia canal in front of the industrial area of Shoubra El Khemia. M. Sc. Thesis, Fac. Sci., Ain Shams Univ., Cairo.
- Abdo, M.H.: 2002, Environmental studies on Rosetta Branch and some chemical applications at the area extends from EL-Kanter EL-Khyria to Kaf EL-Ziate City. Ph.D. Thesis, Fac. Sci., Ain Shams Univ. Cairo.
- Abo Sedera, S.A.: 1990, Studies on microbial pollution in water sources. M. Sc. Thesis, Fac. Agric., El-Azhar Univ. Cairo.
- Ahmed, A. M., Harrington, D.W. and Kane, K.O.: 1995, Control of industrial wastewater in Cairo. Second Middle East Conference on Wastewater Management, Cairo.
- Ali, M.H.H.: 1998, Chemical and physical studies on the River Nile at Damietta branch region. M.Sc. Thesis, Fac. Sci., Menofiya Univ., Shebin El-Kome.
- Anne-Marie P.; Devriese L.A.; Hernandez, J.F. and Delattre, J.M.: 1991, Enumeration by a miniaturized method of *Escherichia coli* and *Streptococcus bovis* as indicators of the origin of faecal pollution of waters. *J. Appl. Bacteriol.* 70: 525-530.
- APHA.: 1998, Standard methods for the examination of water and wastewater. 20th Ed. American Public Health Association, Washington D.C.
- Barnett, H.L.: 1972, Illustrated genera of imperfect fungi. Minneapolis. Burgess Publishing, USA. 241pp.
- Barron, G.L.: 1968, The genera of *Hyphomycetes* from soil. Williams and Wilkins Co., Baltimore.
- Bedair, S.: 2006, Environmental studies on zooplankton and phytoplankton in some polluted areas of the River Nile and their relation with the feeding habit of fish. Ph.D. Thesis, Fac. Sci., Zagazig Univ., Zagazig.
- Bojanic N., Solio, M., Krstulovic, N., Marasovic, I., Nincevic, Z. and Vidjak, O.: 2001, Seasonal and vertical distribution of the ciliated protozoan and micrometazoa in Kastela Bay (Central Adriatic). *Helgoland Marine Res.* Springer-Verlag and Awl.
- Buck, J.D.: 1977, *Candida albicans*. In: Bacterial Indicators, Health Hazards Associated with Water. Hoadley, A.W. and B.J. Dutka (eds.). ASTM Special Technical Publication 635: 139-147.
- Chapman, D.: 1992, Water quality assessments. 1 st ed, Chapman and Hall. London, and New York.
- Cochran, W.G.: 1950, Estimation of bacterial densities by means of the most probable number. *Biometrics* 6 (2): 105-116.
- Dan, T.B.B. and Koppel, F.: 1992, Indicator bacteria for faecal pollution in the littoral zone of Lake Kinneret. *Wat. Res.* 26: 1457-1469.
- Domsch, K.H., Gams, W. and Anderson, T.H.: 1980, Compendium of soil fungi. Acad. Press, London. 859 pp.
- Dutka., B.J.: 1989, Methods for microbiological and toxicological analysis of water, waste water and sediments. Burlington. Canada.
- EL-Bassat, R. A.: 1995, Ecological studies of zooplankton on the River Nile. M.Sc. Thesis, Fac. Sci., Suez Canal Univ., Ismailea.
- EL-Bassat, R.: 2002, Ecological studies of zooplankton communities with particular reference to free living protozoans at River Nile-Damietta Branch .Ph.D. Thesis, Girls College for Arts, Sci. and Educ., Ain Shams Univ., Cairo.
- Elewa, A.A. and Mahdi, H.: 1988, Some limnological studies on the Nile water at Cairo. *Egypt. Bull. Nat. Inst. Oceanogr and Fish.* 14 (2): 141-152.
- Elewa, A.A.: 1991, Influence of flood water on the physical and chemical features of Lake Nubia, Sudan. *Bull. Inst. Oceanogra. and Fish.* 17(1): 97-109.
- Elewa, A.A., Shehata, M.B. and Abdo, M.H.: 2001, Effect of thermal pollution of Shoubra El-Kheima Electric Station on the River Nile water quality. The Second International Conference and Exhibition for Life and Environment, Alexandria: 3-5.
- EL-Sharouny, H.M.M.: 1989, Pollution effects on fungi inhabiting organic debris in the Nile water. *Egypt. J. Microbiol.* 24 (3): 405-414.
- EL-Taweel. G.E.: 1998, Microbiological profile of raw Nile water. *J. Egypt Public Health Assoc.* 73: 5-6.
- Erez, J., Krom, M.D. and Neuwirth, T.: 1990, Daily oxygen variation in marine fishponds. Eilat. *Israel. Aquacul. Goldman Jc Tenore Rk Ryther HJ Corwin* 84: 289-305.
- Ezzat, M.N., Shehab, H., Hassan, A. A., ELSharkawy, M., EL Diasty, A., EL Assiouty, I., EL-Gohary, F. and Tczap, A.: 2002, Survey of Nile system pollution. Report No. 64. Ministry of Resources and Irrigation, US Agency for International Development, Agricultural Policy Reform Program.
- Geldreich, E.E.: 1974, Buffalo lake recreational water quality: a study on bacteriological data interpretation. *Water Res.* 6: 913-921.
- Ghallab, M.H.M.: 2000, Some physical and chemical changes on River Nile down stream of Delta

- Barrage at El-Rahawy drain. M. Sc. Thesis, Fac. Sci, Ain Shams Univ, Cairo.
- Godfree, A.F, Kay, D and Wyer, M.D.: 1997, Faecal streptococci as indicator of faecal contamination in water. *J. Appl. Microbiol.* 83: 1105-1195.
- Hecter, E.M., Danial, E.M., Jose, L.C. and Emilia, B.: 1998, Suburban areas in developing countries and their relationship to ground water pollution: A case study of Mardel plate, Argentina. *Environ. Manag.* 22-245.
- Heikal, M.T.H.: 2000, Environmental studies on antibiotic resistant bacteria in some locations along the River Nile. Ph.D.Thesis, Instit. Environ. Stud. and Res., Ain Shams Univ., Cairo.
- Hesselsøe, A.M.: 2001, Ammonia-oxidizing bacteria and nitrification in soil. Ph. D. Thesis, Royal Veterinary and Agricultural Univ., Denmark.
- Holt, J.G., Krieg, N.R., Sneath, P.H.A., Stalety, J.T. and Williams, S.T.: 1994, Bergey's Manual of Determinative Bacteriology: Facultatively anaerobic gram-negative rods. 9th ed, Williams and Wilkins, Baltimore, London.
- Horna, R.A.: 1972, Marine chemistry. Wiley Interscience, London.
- IOC: 1993, Chlorinated biphenyls in open waters: sampling extraction, clean-up and instrumental determination. Manual and Guides No 27. Inter governmental Oceanographic Commission, UNESCO, Paris.
- Jørgensen, S.K.: 1989, Annual pattern of denitrification and nitrate ammonification in estuarine sediment. *Appl. Environ. Microbiol.* 55 (7): 1841-1847.
- Kizewicz, B.: 2004, *Candida albicans*, *Candida aquatica*, *Candida krusei* and *Candida tropicalis* strains isolated from springs of Podlasie Province. *I: Wiad Parazytol.* 50 (3): 535-544.
- Krom, M.D., Neori, A. and Van Rijn, J.: 1989, Importance of water flow rate in controlling water quality processes in marine and fresh water fishponds. Israel. *J. Aquacul. Goldman Jc Tenore Rk Ryther HJ Corwin* 41: 23-33.
- Lårone, D.H.: 1995, Medically important fungi; a guide to identification, Third Edition, ASM Press, Washington D.C.
- Mahrous, Y.M.: 1997, Sources of River Nile water pollution from Assuit to Cairo and the evaluation of its effect on the water quality. *J. Environ. Sci.* 13: 45-65.
- Martinez- Manzanares, E., Morinigo, M.A., Conax, R., Fgea, F. and Borrego, J.I.: 1991, Relationship between classical indicators and several pathogenic microorganisms involved in shell fish borne diseases. *J. Food Protect.* 54:711-717.
- Naguib, A.I.: 1961, The use of suitable cultural methods for determining the distribution of soil microorganisms under local conditions. U.A.R. Bot. 4-32.
- Niewolak, S., Korycka, A.: 1979, Nitrification processes in fertilized lakes. *Ekol. Pol.* 27: 625-655.
- Noble, R.T. Lee, I.M. and Schiff, K.C.: 2004, Inactivation of indicator microorganisms from various sources of faecal conramination in seawater and fresh water. *J. Appl. Microbiology* 96: 464-472.
- Prinčič, A., Mahne, I., Megušar, F., Paul, A.E, and Tiedje, M.J.: 1998, Effects of pH and oxygen and ammonium concentrations on the community structure of nitrifying bacteria from wastewater. *Applied and Environmental Microbiology* 64 (10): 3584-3590.
- Ramirez, C.: 1982, Manual and atlas of *Penicillia*. Elsevier Biomedical Press, Amsterdam.
- Rabeh, S.A.: 1993, Bacteriological studies on Wadi EL-Raiyan Lake. Faiyum, Egypt. M.Sc. Thesis, Fac. Sci., Univ, Tanta.
- Rabeh, S.A.: 1996, Bacteriological and chemical studies on benthic layers of Wadi El- Raiyan Lake. Faiyum Governorate, A. R. Egypt. Ph. D. Thesis, Fac. Sci, Univ, Tanta.
- Rabeh, S.A.: 2000, Thermal and microbial pollution in the River Nile at the industrial region of Shoubra EL-Kheima. Egypt. *J. Egypt. Acad. Soc. Envir. Develop.* 1(1): 83-98.
- Rabeh, S. A. and Azab, E.: 2006, Bacterial indicators of both sewage pollution and trophic status in Abu Zaabal Lakes. Egypt. *Res. J. Microbiol.* 1: 480-491.
- Rabeh, S.A.: 2007, Monitoring of microbiological and sanitary quality of water in Rosetta Branch of the River Nile, Egypt. *J. Egypt. Acad. Soc. Environ. Develop.* 8 (4): 57-70.
- Rabeh, S.A., EL-Bassat. R.A., Kamel. E.G., Tawfic, T.A. and Amer, A.S.: 2007, The interaction between zooplankton and bacterial indicators of pollution in River Nile at Cairo. Egypt. *African. J. Biol. Sci.* 3 (2): 91-109.
- Raper, K.B. and Fennell, D. I.: 1965, The genus *Aspergillus*. Williams and Wilkins Co., Baltimore.
- Reid, K.G.: 1961, Ecology of inland waters and estuaries. Text book VNR, New York.
- Russo, R.C., Thurston, R.V. and Emerson, K.: 1981, Acute toxicity of nitrite to rainbow trout (*Salmo gairdner*): effects of pH, nitrite species and amino species. *Can. J. Fish. Aquat. Sci.* 38: 387-393.
- Sabae, S.Z. and Rabeh, S.A.: 2007, Evaluation of the microbial quality of the River Nile waters at Damietta Branch, Egypt. *Egypt. J. Aquat. Res.* 33: 301-311.
- Sayed, M.F.: 2002, Chemical studies on pollution in Rosetta branch of River Nile between Kafr El-Zayat and Rosetta outlet. Ph.D. Thesis, Fac. Sci, Cairo Univ. Cairo.
- Shaban, A.M. and EL-Taweel, G.E.: 2003, Microbiological monitoring and evaluation of River Nile water at Cairo segment and Ismailia Canal. *Egyptian Journal of Microbiology* 38 (2): 169-182.
- Siliem, T.A.A.: 1984, Chemical studies on pollution in the Damietta Nile Branch between the Faraskour Dam and Ras EL-Bar outlet. Ph.D. Thesis, Fac. Sci., Alex Univ., Alexandria.

- Uden, N. and Ahearn, D.C.: 1963, Occurrence and population densities of yeast species in a fresh-water lake, *Antonie Van Leeuwenhoek* 29(1): 308-312.
- UNEP/IOC/IAEA: 1991, Sampling of selected marine organisms and sample preparation for the analysis of chlorinated hydrocarbons. Reference Methods for Marine Pollution Studies no 12, Revision 2. Nairobi UNEP. 17.
- Wojcik, A., Rozga, A. and Kurnatowski, P.: 2003, Prevalence of potentially pathogenic fungi in the bathing sites of the Sulejow Reservoir. 1: *Wiad Parazytol.* 49(2): 173-185.

دراسة بيئية ميكروبيولوجية على نهر النيل بمنطقة كفر الزيات الصناعية

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تم دراسة العدد الميكروبي الكلى للفطريات والخمائر والبكتريا لمياه نهر النيل (فرع رشيد) بالقرب من منطقة كفر الزيات الصناعية كمؤشر للتلوث العضوي . وجد أن أعلى قيمة للخمائر كانت في الشتاء أمام مصب شركة الملح و الصودا وعلى بعد كيلو متر واحد منها ($10^2 \times 113$ و $10^2 \times 149$ مستعمرة على التوالي)، وكان *Saccharomyces cerevisiae* و *Kloeckera spp* أكثر الكائنات ظهورا في هذا الموسم. حققت الفطريات أعلى عدد لها خلال الشتاء أمام مصب شركة الملح والصودا أيضا عند درجة حرارة 37 ° (*Trichoderma spp*). ومن الملاحظ أن *Candida albicans* و *Candida magnoliae* كانا أكثر الكائنات ظهورا خلال الربيع والصيف عند درجة حرارة 27 °. سجلت بكتريا القولون الكلية وبكتريا القولون البرازية أعلى عدد لها ($10^3 \times 11$ مستعمرة) في الصيف والشتاء. وقد لوحظت الزيادة الهائلة في العدد الكلى للبكتريا الممرضة خلال موسم الشتاء، وكان من أهم الكائنات التي تم عزلها *E. coli* ، *Sallmonella* ، *Serratia* و *Klebsiella spp*. وتم في هذه الدراسة أيضا تقدير بعض الخواص البيئية للماء مثل درجات الحرارة والتوصيل الكهربى والاس الهيدروجينى. وعموما، تشير كل الخواص الميكروبيولوجية إلى أن فرع رشيد يتعرض للصرف العضوى، ومن خلال الدورالذى تقوم به الخمائر والبكتريا في دورة النيتروجين بمياه النهر، يتضح أن عمل وحدات معالجة ميكروبيولوجية للصرف الصناعي الناتج من الشركات التي تصب مباشرة في نهر النيل ذو فائدة عالية للتغلب نسبيا على مشكلة ارتفاع الامونيا في هذه المياه.