

IONIC REGULATION OF THE BLOOD IN THE CAT FISH CLARIAS LAZERA UNDER THE EFFECT OF EXPERIMENTAL ACID POLLUTION

By

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ABSTRACT

The acute and chronic treated fish showed significant decrease in both Na^+ and Cl^- levels. K^+ levels showed a higher values in the acute case, but an adaptation appears to take place in the chronic case. An increased in the concentrations of Ca^{2+} was observed during both acute and chronic acid exposures.

Na^+ , K^+ and Ca^{2+} ions efflux showed a significant increase 24 hrs following the transference of fish to pH 3.1. A rapid decrease of serum osmolality occurred at both pH levels (5.4 and 4.7), after chronic acid exposure. This decrease was observed throughout the whole of the experimental period. (12 weeks).

INTRODUCTION

Electrolytes are distributed in solution throughout all the body fluids. They have various functions, the most important of which are to contribute a majority of the osmotically active particles, to provide buffer systems and mechanisms for the regulation of pH (acid - base balance), and to give the proper ionic balance for normal neuro - muscular irritability and tissue function (West, 1957). Disturbances in acid - base balance osmoregulation have been emphasized as key toxic mechanisms leading to fish death (Packer., 1979; Graham and Wood, 1981; Mc Donald and Wood, 1981; Booth et al, 1982 and Mc Donald, 1983 b).

The goal of the present study was to investigate the physiological effects of acute and chronic acid stress on the fresh water fish Clarias lazera. Serum Na^+ , K^+ , Ca^{2+} and Cl^- were measured as indices of ionoregulatory disturbances.

MATERIALS AND METHODS

Specimens utilized in the present investigation are freshwater fish *Clarias lazera*, ranging in length from 250 to 300 mm and weighing from 150 to 200 g in weight. They were collected during the summer months June, July and August of the year 1989 and 1990, from Lake Maruit near Alexandria, Egypt. All animals were acclimated for at least two weeks under controlled laboratory conditions. They were fed on commercial dry pellets adlibitum.

Exposure to acid as pure sulphuric acid (H_2SO_4), was carried out using static renewal technique in which test water was periodically replaced, usually once every 24 hr, by a new fresh one of the same composition. Acid concentration to which the fish were subjected was pH 3.1, TL_m pH level for 96 hr exposure and chronic (pH 5.4 and 4.7) for 12 weeks of exposure, in comparison with a control group at pH 7.1.

Blood was sampled for electrolytes assays (by severing the caudal peduncle), allowed to clot and then centrifuged at 8000 for 5 minutes, and serum was stored under refrigeration until the time of analysis.

Serum Na and K were determined by the quantitative analysis of Na and K in serum using ion selective electrode measuring method (Proda and Simon, 1970).

Serum calcium was quantitatively determined using the Cresolphalein complex (CPC) calorimetric method.

Colorimetric determination of chloride ions in serum was undertaken with the TPTZ - mercury complex method (Feld Kamp, 1974).

Determination of the osmotic activity (osmolality) of serum was determined by using the Beckman ASTRA Automated State-Routine Analyzer systems after chronic acid exposure. Comparison of the experimental results with controls was made by student's t-test.

RESULTS

I- In serum samples:

Serum Na concentration was significantly decreased from the control value (105 ± 2.5 mmol / L) to 94.66 ± 0.33 mmol / l ($p < 0.025$) after 24 hours after of exposure to pH 3.1.

The 72 hours exposure showed a nonsignificant decrease, which become significantly lower than the control value at the end of the experiment $93,33 \pm 2.02$ mmol / L ($p < 0.025$). In contrast serum potassium content exhibited a remarkable increase (4.13 ± 0.03 mmol / L) as compared to their corresponding controls (2.26 ± 0.08 mmol / L) after 24 hours exposure which remained significantly higher till the 96 hours period (Table 1)

Chronic exposure of pH 5.4 and 4.7 for 12 weeks caused major changes in serum Na^+ content (Table 2), a marked decrease from the control level (117.0 ± 0.81 mmol / L) was detected starting from the first week of treatment. pH 5.4 caused a drop of the serum Na level to 106.66 ± 1.453 mmol / L ($p < 0.025$) after the 12 weeks of exposure. The effect of lower pH 4.7 was more severe, since the drop of the serum Na^+ level after 12 week of exposure was highly significant reaching the value of 96.33 ± 1.453 mmol / L ($p < 0.001$).

Results for serum K concentrations of fish exposed chronically to pH 5.4 and 4.7 respectively are compared in Table (3) with that of controls. A gradual recovery to the control level was observed in the 11 and 12 week of exposure to pH 5.4 reaching value of 2.75 ± 0.02 and 2.48 ± 0.04 mmol / L comparable to the control values (2.2 ± 0.20 and 2.40 ± 0.12 mmol / L) respectively.

With respect to the pH 4.7 treated group, significant elevations from the control values was observed for 11 weeks of exposure. In the 12 week, only the value obtained was not significant from the control value.

Serum Ca^{2+} concentration of the acute acid treated fish showed a transient decrease from the control level (11.40 ± 0.15 mg / dl) to 10.16 ± 0.12 mg / dl after 24 hours of exposure followed by a significant increase at the 96 hr of exposure (11.9 ± 0.05 mg / dl) $p < 0.05$.

Serum Cl concentration showed a progressive highly significant decrease during the whole experimental period ($p < 0.001$) reaching a level of 57.08 ± 0.61 mmol / l by the 96 hr exposure from the control level of 135.33 ± 0.88 mmol/L. (Table 4).

Table (1): Changes in serum sodium and potassium concentrations juvenile Clarias lazera during exposure to acidified water (pH 3.1) over a 96-h period.

	Exposure period (hours)	Range	Mean± S.E.	S.D.	% Coefficient of variation	Level of significance
Sodium (Na ⁺) (mmol/l)	Control	100-108	105.000±2.517	4.359	4.151	
	24 h	94- 95	94.667±0.333	0.577	0.609	P < 0.025
	48 h	87- 90	88.000±1.000	1.732	1.968	P < 0.005
	72 h	99-102	100.333±0.882	1.527	1.522	N.S
	96 h	90- 97	93.333±2.027	3.512	3.763	P < 0.025
Potassium(K ⁺) (mmol/l)	Control	2.1-2.4	2.267±0.088	0.153	6.739	
	24 h	4.0-4.3	4.133±0.088	0.153	3.696	P < 0.001
	48 h	2.9-3.1	3.000±0.058	0.100	3.333	P < 0.005
	72 h	2.5-2.9	2.700±0.115	0.200	7.407	P < 0.05
	96 h	3.4-3.8	3.633±0.120	0.208	5.729	P < 0.001

- Each mean result corresponds to a mean value of the three experiments.
 S.E. = Standard error.
 S.D. = Standard deviation.

Table (2) Changes in serum sodium concentration (expressed as mmol/l) of juvenile *Clarias lazera* exposed to different treatment with H₂SO₄, pH_s 7.1 (control), 5.4 and 4.7 over a 12-week period.

Exposure period (weeks)	Range	Mean±S.E.	S.D.	%Coefficient of variation	Level of Significance
First week	116-118	117.00±0.816	1.414	1.209	P<0.001 P<0.001
	100.5-104	102.16±1.014	1.756	1.719	
	91-95	93.00±1.155	2.000	2.150	
Second week	112.5-116	114.25±1.429	2.475	2.167	P<0.001 P<0.001
	91.0-94	92.33±0.882	1.527	1.654	
	70.0-78	74.66±2.404	4.163	5.576	
Third week	115-117	116.00±0.816	1.414	1.219	P<0.01 N.S.
	106-110	107.33±1.333	2.309	2.152	
	113-120	117.00±2.082	3.605	3.082	
Fourth week	113-117	115.00±1.633	2.828	2.459	P<0.001 P<0.001
	97-98	97.33±0.333	0.577	0.593	
	89-93	91.00±1.155	2.000	2.198	
Fifth week	113-118	115.50±2.041	3.535	3.061	P<0.001 P<0.001
	91-95	92.66±1.202	2.082	2.246	
	80-87	82.66±2.186	3.786	4.579	
Sixth week	114-120	117.00±2.449	4.243	3.626	P<0.001 P<0.001
	91-94	93.00±1.000	1.732	1.862	
	89-94	91.33±1.453	2.517	2.755	
Seventh week	115-115	115.00±0.000	0.000	0.000	P<0.001 N.S.
	99-103	100.66±1.202	2.082	2.068	
	113-117.5	114.83±1.364	2.363	2.058	
Eighth week	117-118	117.50±0.408	0.707	0.602	N.S. P<0.05
	113-117	115.00±1.155	2.000	1.739	
	115-116	115.66±0.333	0.577	0.499	
Ninth week	116-118	117.00±0.816	1.414	1.209	P<0.001 P<0.01
	96-97	96.66±0.333	0.577	0.597	
	100-109	103.33±2.848	4.933	4.774	
Tenth week	112-114	113.25±0.612	1.061	0.936	P<0.001 P<0.01
	96-98.5	97.33±0.726	1.258	1.293	
	100-107	103.00±2.082	3.605	3.500	
Eleventh week	113.5-115	114.25±0.612	1.061	0.928	P<0.001 P<0.025
	101-103	102.16±0.601	1.041	1.019	
	109.5-122	110.50±0.764	1.323	1.197	
Twelfth week	112-114	113.00±0.816	1.414	1.251	P<0.025 P<0.001
	104-109	106.66±1.453	2.517	2.359	
	94-99	96.33±1.453	2.517	2.612	

- Each mean result corresponds to a mean value of the three experiments.
- pH_s are arranged in the table in a decreasing order (7.1, 5.4, 4.7) within each week.

S.E. = Standard error.

S.D. = Standard deviation.

Table (3): Changes in serum sodium potassium concentration (expressed as mmol/l) of juvenile *Clarias lazera* exposed to different treatment with H₂SO₄, pH, 7.1 (control), 5.4 and 4.7 over a 12-week period.

Exposure period (weeks)	Range	Mean±S.E.	S.D.	% Coefficient of variation	Level of Significance
First week	1.90-2.10	2.000±0.081	0.141	7.071	P<0.001 P<0.001
	4.30-4.40	4.366±0.033	0.058	1.322	
	4.70-4.85	4.783±0.044	0.076	1.597	
Second week	1.85-2.05	1.950±0.081	0.141	7.252	P<0.001 P<0.001
	4.50-4.55	4.516±0.017	0.029	0.639	
	4.80-4.95	4.883±0.044	0.076	1.564	
Third week	2.00-2.10	2.050±0.041	0.071	3.449	P<0.001 P<0.001
	4.65-4.75	4.700±0.029	0.050	1.064	
	4.95-5.05	5.000±0.029	0.050	1.000	
Fourth week	2.00-2.10	2.050±0.041	0.071	3.449	P<0.001 P<0.001
	3.65-3.85	3.783±0.067	0.115	3.052	
	4.35-4.65	4.466±0.093	0.161	3.598	
Fifth week	2.30-2.35	2.325±0.020	0.035	1.521	P<0.001 P<0.001
	3.70-3.75	3.733±0.017	0.029	0.773	
	4.05-4.20	4.117±0.044	0.076	1.855	
Sixth week	2.00-2.20	2.100±0.081	0.141	6.734	P<0.001 P<0.001
	3.95-4.05	4.000±0.029	0.050	1.250	
	4.40-4.65	4.550±0.076	0.132	2.907	
Seventh week	2.55-2.80	2.675±0.102	0.177	6.608	P<0.005 P<0.001
	3.60-3.70	3.717±0.073	0.126	3.385	
	4.00-4.25	4.100±0.076	0.132	3.226	
Eighth week	2.70-2.80	2.750±0.041	0.071	2.571	P<0.001 P<0.001
	3.65-3.80	3.733±0.044	0.076	2.046	
	4.10-4.35	4.200±0.076	0.132	3.149	
Ninth week	2.40-2.65	2.525±0.102	0.177	7.001	P<0.001 P<0.001
	3.50-3.60	3.550±0.029	0.050	1.408	
	3.80-3.90	3.850±0.029	0.050	1.299	
Tenth week	2.40-2.50	2.450±0.041	0.071	2.886	P<0.001 P<0.001
	3.05-3.25	3.133±0.061	0.104	3.322	
	3.60-3.70	3.650±0.029	0.050	1.369	
Eleventh week	1.95-2.45	2.200±0.204	0.353	16.070	N.S. P<0.01
	2.70-2.80	2.750±0.029	0.050	1.818	
	3.25-3.35	3.300±0.029	0.050	1.515	
Twelfth week	2.25-2.55	2.400±0.122	0.212	8.839	N.S. N.S.
	2.40-2.55	2.483±0.044	0.076	3.075	
	2.60-2.90	2.733±0.088	0.153	5.588	

- Each mean result corresponds to a mean value of the three experiments.
 - pH_s are arranged in the table in a decreasing order (7.1, 5.4, 4.7) within each week.

S.E. = Standard error.
 S.D. = Standard deviation.

Table (4): Changes in serum calcium and chloride concentrations of juvenile Clarias lazera during exposure to acidified water (pH 3.1) over a 96-h period.

	Exposure period (hours)	Range	Mean± S.E.	S.D.	% Coefficient of variation	Level of significance
Calcium (Ca ²⁺) (mg/dl)	Control	11.2-11.7	11.400±0.153	0.264	2.321	
	24 h	10.0-10.4	10.167±0.120	0.208	2.047	P < 0.005
	48 h	12.4-12.6	12.500±0.058	0.100	0.800	P < 0.005
	72 h	10.9-11.2	11.067±0.088	0.153	1.380	N.S
	96 h	11.8-12.0	11.900±0.058	0.100	0.840	P < 0.05
Chloride (Cl ⁻) (mmol/l)	Control	134-137	135.333±0.882	1.527	1.129	
	24 h	117-120	118.466±0.667	1.501	1.267	P < 0.001
	48 h	104-106	105.166±0.601	1.041	0.989	P < 0.001
	72 h	80-83.7	82.066±1.089	1.888	2.300	P < 0.001
	96 h	56-58.1	57.080±0.615	1.065	1.866	P < 0.001

- Each mean result corresponds to a mean value of the three experiments.

S.E. = Standard error.

S.D. = Standard deviation.

When compared to control values 11.56 ± 0.032 , changes induced in Ca^+ ion concentration, due to chronic exposure of fish to acid, were significantly elevated throughout the duration of experiment (12 weeks) for both pH 5.4 (11.887 ± 0.009 , $p < 0.001$) and pH 4.7 (12.060 ± 0.046 , $p < 0.001$). Table (5).

With respect to chloride ion levels, significant decreasing values were observed throughout the chronic pH 5.4 exposure period.

12 week after exposure the Cl ion concentration decreased to 77.706 ± 0.633 with compared to control value 102.055 ± 0.066 . At 4.7, the loss of Cl ions was somewhat more pronounced which start and remained highly significant ($p < 0.001$) from the corresponding control values till the end of the experiment. Table (6). A dramatic decrease amounting to about 50 % loss was recorded during the last week of exposure giving a value of 54.46 ± 0.45 mmol/L.

II- In water samples:

Determination of whole body sodium, potassium and calcium ion efflux.

Flame photometric analysis of water electrolyte were performed on acid exposed (pH 3.1) and control (pH 7.1) fish to a certain that an electrolyte in balance is associated with acid exposure. were illustrated in Table 7 Sodium ion efflux was observed within 60 minutes showing a significant level of $p < 0.05$. A gradual continual efflux, ending with a highly significant maximum value of 65.04 ± 0.18 ppm ($p < 0.001$) when compared to control value (59.43 ± 0.08 ppm) was observed.

Potassium and calcium ions efflux showed a time dependent increase following the introduction of fish to pH 3.1 being significant only after 24 hours for potassium reaching a value of 10.86 ± 0.80 ppm ($p < 0.05$), comparable to the control value (8.18 ± 0.44 ppm), and after 45 minutes for calcium reaching a level of 64.00 ± 0.16 ppm ($p < 0.025$) from the untreated control level (61.66 ± 0.53 ppm).

Determination of the osmotic activity (osmolality)

Major changes in the osmolality levels of fish at both pH levels (5.4 and 4.7) occurred after chronic acid exposure as shown in table (Table 8). A substantial and rapid decrease was observed starting from the first week after treatment, recording values of 236.78 ± 0.949 and 22.05 ± 1.114 milliosmoles / kg for pH, 5.4 and 4.7 at the end of the experimental period (12 weeks), compared to their corresponding control value (272.77 ± 3.45 milliosmoles / kg).

Table (5) Changes in serum calcium concentration (expressed as mg/dl) of juvenile *Clarias lazera* exposed to different treatment with H₂SO₄, pH_s 7.1 (control), 5.4 and 4.7 over a 12-week period.

Exposure period (weeks)	Range	Mean±S.E.	S.D.	%Coefficient of variation	Level of Significance
First week	11.40-11.60	11.500±0.081	0.141	1.229	P<0.025 P<0.001
	11.80-11.89	11.843±0.026	0.045	0.381	
	12.48-12.60	12.530±0.036	0.062	0.498	
Second week	11.40-11.50	11.450±0.041	0.071	0.617	P<0.001 P<0.001
	12.04-12.12	12.086±0.024	0.042	0.344	
	13.69-13.72	13.703±0.009	0.015	0.115	
Third week	11.10-11.30	11.200±0.081	0.141	1.263	P<0.001 P<0.001
	12.81-12.89	12.853±0.023	0.040	0.314	
	14.02-14.13	14.083±0.033	0.057	0.404	
Fourth week	10.90-11.00	10.950±0.041	0.071	0.646	P<0.001 P<0.001
	12.89-12.96	12.917±0.022	0.038	0.293	
	13.84-13.91	13.873±0.020	0.035	0.253	
Fifth week	10.80-11.30	11.050±0.204	0.353	3.199	P<0.025 P<0.001
	11.89-11.98	11.936±0.026	0.045	0.038	
	13.28-13.32	13.300±0.011	0.020	0.150	
Sixth week	11.54-11.60	11.570±0.024	0.042	0.367	N.S. P<0.001
	11.47-11.57	11.520±0.029	0.050	0.434	
	12.01-12.09	12.043±0.024	0.042	0.346	
Seventh week	11.41-11.56	11.485±0.061	0.106	0.923	N.S. P<0.001
	11.54-11.60	11.577±0.018	0.032	0.278	
	12.30-12.44	12.373±0.040	0.070	0.566	
Eighth week	11.60-11.70	11.650±0.040	0.070	0.607	P<0.01 P<0.001
	11.87-11.96	11.913±0.026	0.045	0.378	
	12.86-12.94	12.890±0.023	0.040	0.313	
Ninth week	11.85-11.90	11.875±0.020	0.035	0.298	P<0.001 P<0.001
	12.10-12.17	12.137±0.020	0.035	0.289	
	12.89-12.93	12.906±0.012	0.021	0.161	
Tenth week	11.30-11.46	11.380±0.065	0.113	0.994	P<0.001 P<0.001
	12.26-12.34	12.303±0.023	0.040	0.328	
	12.87-12.98	12.930±0.032	0.056	0.431	
Eleventh week	11.26-11.38	11.320±0.049	0.085	0.749	P<0.001 P<0.001
	11.94-12.00	11.966±0.018	0.030	0.255	
	12.59-12.69	12.640±0.050	0.398	0.029	
Twelfth week	11.52-11.60	11.560±0.032	0.056	0.489	P<0.001 P<0.001
	11.87-11.90	11.887±0.009	0.015	0.128	
	12.00-12.15	12.060±0.046	0.079	0.658	

- Each mean result corresponds to a mean value of the three experiments.
- pH_s are arranged in the table in a decreasing order (7.1, 5.4, 4.7) within each week.

S.E. = Standard error.

Table (6): Changes in serum chloride concentration (expressed as mmol/l) of juvenile *Clarias lazera* exposed to different treatment with H₂SO₄, pH_s 7.1 (control), 5.4 and 4.7 over a 12-week period.

Exposure period (weeks)	Range	Mean±S.E.	S.D.	%Coefficient of variation	Level of Significance
First week	100-104	102.000±1.633	2.828	2.773	P<0.005 P<0.001
	88-90	89.203±0.612	1.602	1.188	
	75-79	77.000±1.154	2.000	2.597	
Second week	100-103	101.500±1.224	2.121	2.089	P<0.001 P<0.001
	87-89.1	87.780±0.664	1.149	1.309	
	74-76	75.320±0.663	1.149	1.525	
Third week	99.60-100.13	99.865±0.216	0.375	0.375	P<0.001 P<0.001
	89.90-91.40	90.520±0.452	0.783	0.865	
	73.00-73.90	73.510±0.236	0.462	0.628	
Fourth week	94.90-97.10	96.000±0.898	1.556	1.620	N.S. P<0.001
	93.81-96.03	95.160±0.684	1.185	1.245	
	70.84-73.46	72.013±0.768	1.331	1.848	
Fifth week	95.00-97.00	96.000±0.816	1.414	1.473	N.S. P<0.001
	93.22-99.32	96.410±1.766	3.059	3.173	
	78.89-81.04	80.040±0.625	1.083	1.354	
Sixth week	99.85-100.00	99.925±0.061	0.106	0.106	N.S. P<0.001
	99.75-100.10	99.970±0.111	0.197	0.192	
	87.54-88.22	87.960±0.213	0.369	0.419	
Seventh week	101.00-101.19	101.095±0.077	0.134	0.133	P<0.001 P<0.001
	85.67-88.50	86.780±0.871	1.508	1.738	
	74.13-75.40	74.870±0.383	0.664	0.886	
Eighth week	102.11-102.71	102.410±0.245	0.424	0.414	P<0.001 P<0.001
	80.70-84.74	82.610±1.171	2.028	2.455	
	61.61-64.89	62.980±0.984	1.705	2.707	
Ninth week	102.76-103.35	103.055±0.241	0.417	0.405	P<0.001 P<0.001
	81.13-85.20	83.640±1.268	2.197	2.626	
	60.64-60.91	60.800±0.083	0.144	0.236	
Tenth week	100.59-101.69	101.140±0.202	0.778	0.769	P<0.001 P<0.001
	82.00-83.03	82.490±0.298	0.516	0.625	
	58.65-59.74	59.136±0.320	0.554	0.937	
Eleventh week	100.18-100.59	100.385±0.028	0.289	0.289	P<0.001 P<0.001
	79.91-81.62	80.510±0.554	0.959	1.192	
	54.20-55.11	54.540±0.287	0.497	0.911	
Twelfth week	101.74-102.37	102.055±0.066	0.445	0.436	P<0.001 P<0.001
	76.99-78.97	77.706±0.633	1.097	1.412	
	53.55-54.96	54.460±0.459	0.795	1.458	

- Each mean result corresponds to a mean value of the three experiments.
 - pH_s are arranged in the table in a decreasing order (7.1, 5.4, 4.7) within each week.

S.E. = Standard error.

S.D. = Standard deviation.

Table (7): Changes in whole body sodium, potassium and calcium ions efflux (expressed as ppm) of juvenile *Clarias lazera* exposed to acidified water (pH 3.1, TSM value) over a 24-h period.

Exposure Time	Ion efflux values							
	Sodium (Na ⁺)		Potassium (K ⁺)		Calcium (Ca ²⁺)			
	Mean control values ± S.E	Mean treated values ± S.E	Mean control values ± S.E.	Mean treated values ± S.E.	Mean control values ± S.E.	Mean treated values ± S.E.		
0 min	56.016±0.140	56.178±0.142	6.535±0.112	6.535±0.794	60.833±0.772	60.833±0.224		
15 min	56.097±0.092	56.747±1.010	6.614±0.197	6850±0.894	60.833±0.736	62.005±0.335		
30 min	56.910±0.211	57.723±0.968	6.850±0.221	7.165±0.883	61.666±0.711	62.055±0.087		
45 min	57.154±0.134	59.105±0.911	7.086±0.168	7.244±0.646	61.666±0.534	64.000±0.169		
60 min	57.723±0.088	59.593±0.584 ^a	7.322±0.089	7.480±0.872	66.666±0.421	66.666±0.156		
2 h	58.048±0.110	61.544±0.666 ^b	7.559±0.342	7.559±0.967	66.666±0.411	66.666±0.311		
3 h	58.861±0.243	61.788±0.789 ^a	7.559±0.021	8.031±0.873	67.005±0.672	68.333±0.071		
4 h	59.186±0.097	62.195±0.594 ^b	7.874±0.167	8.031±0.734	68.333±0.495	70.833±0.376		
24 h	59.430±0.081	65.040±0.183 ^b	8.188±0.442	10.866±0.801 ^a	70.833±0.663	74.166±0.258		

- Each result corresponds to a mean value of the three experiments.

S.E. = Standard error.

- Control pH = 7.1 ± 0.1.

Table (8): Changes in serum osmoticactivity (osmolality) (expressed as milliosmoles/Kg) ofjuvenile Clarias lazera exposed to different treatment with H₂SO₄, pH_s 7.1 (control), 5.4 and 4.7 over a 12-week period.

Exposure period (weeks)	Range	Mean±S.E.	S.D.	% Coefficient of variation	Level of Significance
First week	268.54-277.00	272.770±3.454	5.982	2.193	P<0.001 P<0.001
	240.72-242.01	241.313±0.376	0.651	0.269	
	232.32-237.34	235.050±1.466	2.539	1.080	
Second week	267.02-268.60	267.810±0.645	1.117	0.417	P<0.001 P<0.001
	237.00-239.04	237.727±0.658	1.139	0.479	
	200.94-205.44	202.513±1.465	2.537	1.253	
Third week	250.67-258.66	254.665±3.261	5.649	2.218	P<0.005 P<0.001
	230.06-234.12	231.807±1.206	2.088	0.901	
	168.31-169.58	168.867±0.375	0.649	0.384	
Fourth week	267.86-273.38	270.620±2.253	3.903	1.442	P<0.001 P<0.001
	219.86-222.14	221.243±0.702	1.215	0.549	
	155.44-164.94	159.490±2.830	4.902	3.074	
Fifth week	268.13-271.60	269.865±1.417	2.454	0.909	P<0.001 P<0.001
	251.60-253.53	252.796±0.603	1.045	0.413	
	221.18-231.10	226.506±2.887	5.005	2.208	
Sixth week	264.27-268.54	266.405±1.743	3.019	1.133	P<0.025 P<0.001
	257.94-259.18	258.693±0.382	0.662	0.256	
	210.18-216.27	213.587±1.795	3.109	1.455	
Seventh week	279.33-290.14	284.735±4.413	7.644	2.684	P<0.025 P<0.001
	263.53-268.20	265.833±1.348	2.336	0.879	
	230.96-235.02	233.093±1.176	2.038	0.874	
Eighth week	250.67-254.04	252.350±1.376	2.383	0.944	P<0.025 P<0.025
	258.20-262.82	260.233±1.362	2.359	0.906	
	243.38-247.80	245.366±1.295	2.243	0.914	
Ninth week	273.38-277.40	275.390±1.641	2.842	1.032	P<0.001 P<0.001
	248.71-251.18	249.660±0.765	1.326	0.531	
	237.08-225.98	238.783±0.891	1.544	0.646	
Tenth week	262.35-265.35	263.850±1.224	2.121	0.804	P<0.001 P<0.001
	240.00-243.18	241.966±0.992	1.719	0.710	
	223.67-225.98	224.606±0.702	1.215	0.541	
Eleventh week	261.77-262.82	262.295±0.428	0.742	0.283	P<0.001 P<0.001
	239.18-243.81	241.473±1.337	2.315	0.959	
	220.70-224.68	222.440±1.176	2.036	0.905	
Twelfth week	252.17-255.22	253.695±1.245	2.157	0.850	P<0.001 P<0.001
	234.94-238.17	236.786±0.949	1.643	0.694	
	220.60-224.24	222.050±1.114	1.929	0.869	

- Each mean result corresponds to a mean value of the three experiments.
 - pH_s are arranged in the table in a decreasing order (7.1, 5.4, 4.7) within each week.

S.E. = Standard error.

S.D. = Standard deviation.

DISCUSSION

Exposure of Clarias lazera in the present study to acute acid stress, led to a pronounced drop in serum Na^+ and Cl^- levels. Serum Na^+ and Cl^- depletion as a result of acute exposure has been demonstrated for a variety of species such as yellow Perch flavescens (Lyons, 1982), large scale sucker Catostomus macrocheilus (McKeown et al, 1985), brook trout Salvelinus fontinalis (Packer and Dunson, 1970), rainbow trout salmon Salmo salar (McWilliams, 1980 b, and Booth et. al., 1982), juvenile rainbow trout (Barton et. al., 1985) and juvenile salmonids (Weiner, 1984 and Wood et al, 1988 a&b).

The decline in serum Na^+ and Cl^- concentrations were detected after exposure of Clarias lazera to chronic acid stress (pH, 5.4 and 4.7). This is in agreement with a previous studies by Leivestad, et al. (1976) and Mc Williams (1980 b).

The following study also indicates that the lower pH 4.7 appears to have a greater significant effect on both electrolytes than the much higher pH 5.4 level (Mc Donald et al., 1983 a, and Packer and Dunson, 1972).

There have been a number of previous studies on the chronic ion regulatory effects (2 weeks to 3 months) of low pH exposure (Mc Williams, 1980 a ; Lee et. al., 1983; Brown et. al., 1986 b; Weiner et. al., 1986; Jones et. al., 1987 and Audet et. al., 1988).

The reduced uptake of Na^+ in acid waters was explained to be due to an increased load on the Na^+ pump mechanism as a consequence serum ionic levels were decreased. Cl^- loss during acute and chronic acid exposure may be a result of passive Cl^- efflux that exceeded active uptake.

Increased serum K^+ resulting from exposure to environmental acidification has been observed for rainbow trout at pH₄ (Mc Donald et. al., 1980 & Booth et. al. 1982, and for carp Cyprinus carpio at pH₄, but not at pH 5.1 (Ultsch et. al., 1981) and Wood et. al., 1988 b).

Clarias lazera exposed to low pH of 5.4 and 4.7 during chronic experiments showed a significant increase in serum K^+ level, followed by a gradual recovery to the control at the end of the experimental period. Rainbow trout Salmo gairdneri exposed to low pH 4.9 and 5.6 (McKeown et. al., 1985) behaved somewhat similar to Clarias lazera. Experiments of Leivestad et. al., 1976 and Mc Donald and Wood, 1981) reported an increased loss of K^+ from muscle tissue at low pH, which might account for serum increases in K^+ . A recovery showed during the chronic experiment may be due to that fish were partially restoring the ion level by

time either by partial reduction of extracellular space, breakdown of erythrocytes thus releasing K^+ or releasing K^+ from cells buffering H^+ to maintain normal cell's electrical potential.

The present data indicate a slight but significant increase in serum calcium levels during acute exposure. This was in accordance with data obtained using Oreochromis mossambicus exposed to pH 4, Wendelaar Bonga *et. al.*, (1984) related this elevation to the indirect effect of the enhanced prolactin secretion resulting in a significant hypercalcemia that was also observed after 2 weeks of acid exposure. In Tilapia, prolactin stimulates the active uptake of Calcium through the gills and has a hypercalcemic action (Wendelaar Bonga and Flik, 1982 and Wendelaar Bonga *et. al.*, 1984).

The increase of calcium level caused by chronic exposure to low pH_s (5.4 and 4.7) in the present study, has been previously observed in other fish species (Holmes and Donaldson, 1969; Whitehead *et. al.*, 1978). Saunders *et. al.*, (1983) who observed high calcium level in Salmon Salmo salar reared at low pH during much of the experiment (5 months), and suggested that this elevation may be an indication of calcium mobilization from skeletal compartments, impaired renal excretion of calcium, elevated calcium uptake from brachial or gut regions, or some combination of these mechanisms. However, it was not clear from the study which, if any, of these mechanisms was responsible for the elevated calcium level.

In the present investigation, acute acid exposure (pH 3.1) for 24 hour was observed to provoke major alterations in ionic balance in Clarias lazera.

Along list of studies (Packer and Dunson, 1970 and 1972; Maetz, 1973; Ashcom, 1979; McWilliams, 1982b; McDonald, 1983 b; Hobe *et al.*; 1984a, Audet and Wood, 1988; Wood 1988 and Wood *et al.*, 1990 showed that exposure of fish to external acidic pH results in an inhibition of active sodium influx and a stimulation of its diffusional efflux, leading to net sodium loss and reduction of body Na levels.

With regards to potassium ion concentration in acutely exposed Clarias lazera to acid 3.1, although serum K concentration was significantly increased after 24 hours, potassium efflux was also found to increase during the efflux experiments, but values were significantly higher only at the final sample (24 hours). These findings suggest that Clarias lazera was capable of either maintaining or restoring serum potassium concentration if only on this short term faces, through some adaptive mechanism. Similar results were demonstrated previously by Booth *et. al.*, 1982.

With respect to the calcium ion reported in the present study on Clarias lazera exposed to pH 3.1, initially calcium efflux was nonsignificant compared to their corresponding controls, until 45 minutes of exposure where Ca efflux became significantly different from controls.

Low pH (4.8) treatment for 24 hours also stimulated Ca efflux in rainbow trout Salmo gairdneri (Reid and McDonald, 1988). Such changes in Ca permeability may also account for most of the drop in plasma Ca following acute exposure.

Several mechanisms have been proposed to explain the effect of H⁺ ion on ionic efflux, but most are encompassed by increased brachial permeability to water and ions (McDonald, 1983 -b, Lee *et. al.*, 1983; and Fryer *et. al.*, 1988).

At both pH, 5.4 and 4.7 in the present chronic exposure systems, serumosmolality dropped abruptly during the first four weeks. The reduction was more severe at pH 4.7. After 4 weeks a gradual increase followed in both pH groups which again showed progressive decreases after 7 weeks at pH 5.4 and 9 weeks at pH 4.7 until termination of the experiment. The initial drop and successive return of osmolality level has been repeatedly reported in different fish species, for different durations (14 - 52 days) and at different pH level (4.5 - 5.5)(Giles *et. al.*, 1984; McKeown *et. al.*, 1985; Brown *et. al.*, 1986 b; Jones *et. al.*, 1987; Audet *et. al.*, 1988 and Fryer *et. al.*, 1988).

The present data obtained on serum osmolality level in Clarias lazera chronically exposed to low pH reinforce the view that reduced pH enhances that ion loss that fish in freshwater typically experience, and that osmolality as a related parameter provide some additional insight into the significance of the increased efflux of ions due to gill permeability changes.

REFERENCES

- Ashcom, T. L. (1979). Serum cortisol and electrolyte responses in acid - stressed brook trout (Salvelinus fontinalis). Ph. D. thesis, Pennsylvania State University.
- Audet, C., Munger, R. S., and Wood, C. M. (1988). Long - term sublethal acid exposure in rainbow trout (Salmo gairdneri) in soft water: effects on ion exchanges and blood chemistry. *Can. J. Fish. Aquat. Sci.* 45: 1387-1398.
- Audet, C., and Wood, C. M. (1988). Do rainbow trout (Salmo gairdneri) acclimate to low pH. *Can. J. Fish. Aquat. Sci.* 45: 1399-1405.

- Barton, B. A., Weiner, G.S., and Schreck, C. B. (1985). Effect of prior acid exposure on physiological responses of juvenile rainbow trout (*Salmo gairdneri*) to acute handline stress. *Can. J. Fish. Aquat. Sci.* 42: 710-717.
- Booth, J. H., Jansz, G. F., and Holeton, G. F. (1982). Cl⁻, K⁺, and acid-base balance in rainbow trout during exposure to, and recovery from, sublethal environmental acidification. *Can. J. Zool.* 60: 1123-1130.
- Brown, S. B., Evans, R. E., and Hara, T. J. (1986b). Internal, thyroidal, carbohydrate, and electrolyte responses in rainbow trout (*Salmo gairdneri*) during recovery from the effects of acidification. *Can. J. Fish. Aquat. Sci.* 43: 714-718.
- Feldkamp, C.S.; 1974. *Z. Klin. Chem. Klin. Biochem.* 12: 146-1500.
- Fried, R., 1972. *Z. Klin. Chem. Klin. Biochem.* 10: 280.
- Fryer, J. N., Tam, W. H., Valentine, B. and Tikkala, R. E. (1988). Prolactin cell cytology, plasma electrolytes, and whole-body sodium efflux in acid stressed brook trout (*Salvelinus fontinalis*). *Can. J. Fish. Aquat. Sci.* 45: 1212-1221.
- Giles, M. A., Majewski, H. S., and Horden, B. (1984). Osmoregulatory and hematological responses of rainbow trout (*Salmo gairdneri*) to extended environmental acidifications. *Can. J. Fish. Aquat. Sci.* 41: 1686-1694.
- Graham, M. S., and Wood, C. M. (1981). Toxicity of environmental acid to the rainbow trout: interactions of water hardness, acid type, and exercise. *Can. J. Zool.* 59: 1518-1526.
- Hobe, H., Laurent, P., and McMahan, B. R. (1984a). Whole body calcium flux rates in freshwater teleosts as a function of ambient calcium and pH levels: a comparison between the euryhaline trout (*Salmo gairdneri*) and the stenohaline bull head (*Ictalurus nebulosus*). *J. Exp. Biol.* 113: 237-252.
- Holmes, W. N., and Donaldson, E. M. (1969). The body compartments and the distribution of electrolytes. In *Fish physiology*. Vol. 1. Edited by W. S. Hoar and D. J. Randall. Academic Press, New York, N. Y. pp. 1-89.
- Jones, K. A., Brown, S. B., and Hara, T. J. (1987). Behavioral and biochemical studies of onset and recovery from acid stress in Arctic charr (*Salvelinus alpinus*). *Can. J. Fish. Aquat. Sci.* 44: 373-381.

- Lee, R. M., Gerking, S. D., and Jezierska, B. (1983). Electrolyte balance and energy metabolism in acid-stressed rainbow trout (*Salmo gairdneri*) and their relation to reproductive success. *Environ. Biol. Fishes.* 8: 115-123.
- Leivestad, H., Hendrey, G., Muniz, I.P., and Snekvik, E., 1976. Effects of acid precipitation on freshwater organisms. SNSF - report 6: Impact of acid precipitation on forest and freshwater ecosystem in Norway Edited by Breakke, F.) pp. 87-111.
- Lyons, J. (1982). Effects of lethal acidity on plasma sodium concentrations in yellow perch (*Perca flavescens*) from a naturally acidic and a naturally alkaline lake. *Comp. Biochem. Physiol.* 73A: 437-440.
- Maetz, J. (1973). $\text{Na}^+/\text{NH}_4^+$, Na^+/H^+ exchanges and NH_3 movement across the gill of (*Carassius auratus*). *J. Exp. Biol.* 58: 255-275.
- McDonald, D. G. (1983b). The interaction of environmental calcium and low pH on the physiology of the rainbow trout (*Salmo gairdneri*). I. Branchial and renal net ion and H^+ fluxes. *J. Exp. Biol.* 102: 123-140.
- McDonald, D. G., Hobe, H., and Wood, C. M. (1980). The influence of calcium on the physiological responses of the rainbow trout (*Salmo gairdneri*) to low environmental pH. *J. Exp. Biol.* 88: 109-131.
- McDonald, D. G., Walker, R. L., and Wilkes, P. R. H. (1983). The interaction of environmental calcium and low pH on the physiology of the rainbowtrout (*Salmo gairdneri*). II. Branchial ionoregulation mechanisms. *J. Exp. Biol.* 102: 141-155.
- McDonald, D. G., and Wood, C. M. (1981). Branchial and renal ion fluxes in the rainbow trout (*Salmo gairdenri*) at low environmental pH. *J. Exp. Biol.* 93: 101-118.
- McKeown, B. A., Geen, G. H., Watson, T. A., Powell, J. F., and Parker, D. B. (1985). The effect of pH on plasma electrolytes, carbonic anhydrase and ATPase activities in rainbow trout (*Salmo gairdneri*) and largescale suckers (*Catostomus macrocheilus*). *Comp. Biochem. Physiol.* 80A: 507-514.
- McWilliams, P. G. (1980a). Acclimation to an acid medium in the brown trout (*Salmo trutta*). *J. Exp. Biol.* 88: 269-280.
- McWilliams, P.G., 1980b. Effects of pH on sodium uptake in Norwegian brown trout (*Salmo trutta*, from an acid river. *J. Exp. Biol.* 88: 259-267.
- McWilliams, P. G. (1982b). The effects of calcium on sodium fluxes in the brown trout (*Salmo trutta*) in neutral and acid water. *J. Exp. Biol.* 96: 439-442.

- Packer, R. K. (1979). Acid-base balance and gas exchange in brook trout (*Salvelinus fontinalis*) exposed to acidic environments. *J. Exp. Biol.* 79: 127-134.
- Packer, R. K., and Dunson, W. A. (1970). Effects of low environmental pH on blood pH and sodium balance of brook trout. *J. Exp. Zool.* 174: 65-72.
- Packer, R. K., and Dunson, W. A. (1972). Anoxia and sodium loos associated with the death of brook trout at low pH. *Comp. Biochem. Physiol.* 41A: 17-26.
- Proda, L., and Simon, W. (1970). Determination of Potassium concentration of serum using a highly selective liquid membrane electrode. *Clin. Acta.* 29: 289-293.
- Reid, S. D., and McDonald, D. G. (1988). Effects of cadmium, copper, and low pH on ion fluxes in the rainbow trout, (*Salmo gairdneri*). *Can. J. Fish. Aquat. Sci.* 45: 244-253.
- Saunders, R.L., Henderson, E.B., P.R., Johnston, C.E. and Fales. J.G., 1983. Effects of low environmental pH on smolting Atlantic Salmon (*Salmosalar*). *Can. J. Fish. Aquat. Sci.* 40: 1203-1211.
- Ultsch, G. R., Ott, M. E., and Heisler, N. (1981). Acid-base and electrolyte status in carp (*Cyprinus carpio*) exposed to low environmental pH. *J. Exp. Biol.* 93: 65-80.
- Weiner, G. S. (1984). Influences of environmental acidification on salmonid reproduction. M. S. thesis, Oregon State University, Corvallis, OR. 40p.
- Weiner, G. S., Schreck, C. B., and Hiram, W. L. (1986). Effects of low pH on reproduction of rainbow trout. *Trans. Am. Fish. Soc.* 115: 75-82.
- Wendelaar Bonga, S.E., and Filk, G., 1982. Prolactin and calcium metabolism in teleost fish. In comparative Endocrinology of Calcium Regulation (C. oguro and PK.T Pange, eds). pp. 21-26 Japan Scientific Press, Tokyo.
- Wendelaar Bonga, S. E., Van Der Meij. J. C. A., and Flik, G. (1984). Prolactin and acid stress in the teleost (*Oreochromis*) formerly (*Sarotherodon mossambicus*). *Gen. Comp. Endocrinol.* 55: 323-332.
- West, E. S. (1957). Text book of biophysical chemistry 2nd ed. The Macmillan company, N. Y. 485p.
- Whitehead, C., Bromage, N. R., and Forster, J. R. M. (1978). Seasonal changes in reproductive function of the rainbow trout (*Salmo gairdneri*). *J. Fish Biol.* 12: 601-608.

- Witters, H. E., and Vandenberght, O. (ED.). (1987). Ecophysiology of acid stress in aquatic organisms. *Anal. Soc. R. Zool. Belg.* 117 (Suppl.1) 472p.
- Wood, C. M. (1988). The physiological problems of fish in acid waters. In R. Morris, D. J. A. Brown, E. W. Taylor and J. A. Brown [ed.] *Acid toxicity and aquatic animals. Society for Experimental Biology Seminar Series*, Cambridge University Press, Cambridge.
- Wood, C. M., McDonald, D. G., Booth, C. E., Simons, B. P., Ingersoll, C. G., and Bergman, H. L. (1988a). Physiological evidence of acclimation to acid/aluminum stress in adult brook trout (*Salvelinus fontinalis*). 1. Blood composition and net sodium fluxes. *Can. J. Fish. Aquat. Sci.* 45: 1587-1596.
- Wood, C. M., McDonald, D. G., Ingersoll, C. G., Mount, D. R., Johannsson, O. E., Landsberger, S., and Bergman, H. L. (1990). Effects of water acidity, calcium, and aluminum on whole body ions of brook trout (*Salvelinus fontinalis*) continuously exposed from fertilization to swim-up: a study by instrumental neutron activation analysis. *Can. J. Fish. Aquat. Sci.* 47: 1593-1603.
- Wood, C. M., Playle, R. C., Simons, B. P., Goss, G. G., and McDonald, D. G. (1988b). Blood gases, acid-base status, ions, and haematology in adult brook trout (*Salvelinus fontinalis*) under acid/aluminum exposure. *Can. J. Fish. Aquat. Sci.* 45: 1575-1586.